

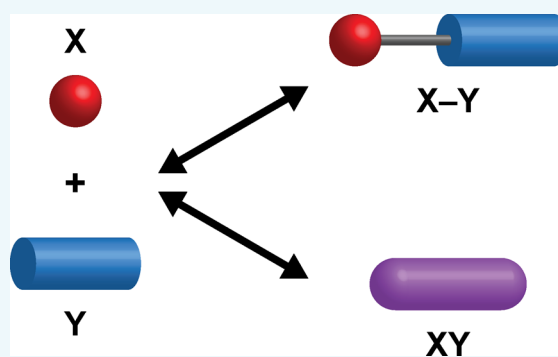
# What Can Pleiotropic Proteins in Innate Immunity Teach Us about Bioconjugation and Molecular Design?

Michelle W. Lee,<sup>†</sup> Ernest Y. Lee,<sup>†</sup> and Gerard C. L. Wong<sup>\*,†,‡,#</sup>

<sup>†</sup>Department of Bioengineering, <sup>‡</sup>Department of Chemistry & Biochemistry, and <sup>#</sup>California NanoSystems Institute, University of California, Los Angeles, Los Angeles, California 90095, United States

## Supporting Information

**ABSTRACT:** A common bioengineering strategy to add function to a given molecule is by conjugation of a new moiety onto that molecule. Adding multiple functions in this way becomes increasingly challenging and leads to composite molecules with larger molecular weights. In this review, we attempt to gain a new perspective by looking at this problem in reverse, by examining nature's strategies of multiplexing different functions into the same pleiotropic molecule using emerging analysis techniques such as machine learning. We concentrate on examples from the innate immune system, which employs a finite repertoire of molecules for a broad range of tasks. An improved understanding of how diverse functions are multiplexed into a single molecule can inspire new approaches for the deterministic design of multifunctional molecules.



## 1. INTRODUCTION

Synthetically modified molecules are known to have a broad range of functions, from targeted drug delivery to biomarker imaging to tracking of intracellular events.<sup>1</sup> A common strategy to add function to a given molecule is by conjugation of a new moiety onto that molecule. As an index of the impressive progress, the repertoire of conjugation techniques used to attach synthetic chemical groups to proteins has grown drastically in recent years.<sup>2</sup> As with all fields, there exist both contingent and structural difficulties. For instance, aside from the chemistry of bioconjugation, care must usually be taken to ensure that there is minimal interference between different parts of the resultant composite molecule. Moreover, adding multiple functions in this way leads to progressively more difficult chemistry and larger molecular weights. In this review, we attempt to gain a new perspective by looking at this problem in reverse, by examining so-called pleiotropic proteins, proteins that have multiple functions encoded into the same structure.

Over the past few decades, multifunctionality has been discovered in an increasing number of natural proteins. These so-called pleiotropic proteins with multiple coexisting roles can be considered products of natural bioconjugation, through which the introduction of covalent structural changes confers new functions.<sup>3</sup> Due to the prevalence of proteins simultaneously involved in several cellular processes, multifunctionality is now believed to be the rule rather than the exception. In fact, over 60% of the proteins in archaea and bacteria, and over 80% of eukaryotic proteins, contain more than one functional domain.<sup>4</sup> In addition, hundreds of “moonlighting” proteins, a subclass of multifunctional proteins, have been identified to

perform two or more distinct functions within a single domain and are not the result of gene fusions, alternative splicing, or multiple proteolytic fragments.<sup>5–7</sup> One example is phosphoglucose isomerase, which is a cytosolic glycolytic enzyme involved in energy metabolism. When secreted from cells, it also plays a dual role as the extracellular cytokine neuroleukin.<sup>6</sup> While incorporating multiple functions into one protein may be nature's method of doing more with less, protein multifunctionality introduces additional levels of complexity that can complicate efforts in understanding physiological processes and molecular mechanisms of disease.

With the significant implications of protein multifunctionality for human health, in both normal and pathological contexts, expanding on our current knowledge of multifunctional proteins can aid in the identification of other proteins and protein families that may also harbor additional functions, and provide insight on how these additional functions evolve. Furthermore, an improved understanding of how multifunctional proteins orchestrate their diverse roles also can enable the design and development of new therapeutics that can perform a multitude of actions, as well as more accurately target the specific roles of multifunctional proteins.

In this review, we present examples from the innate immune system, where multifunctionality is a matter of survival, given that a finite repertoire of molecules must organize against diverse and often unanticipated threats. Specifically, as a baseline example, we will start with a distinct function that

Received: March 8, 2018

Revised: May 10, 2018

Published: May 17, 2018

seems simple enough, that of membrane remodeling and permeation. The point of the review is not to offer an exposition of current ideas on membrane remodeling or attempt a definitive and comprehensive account of the historical development of these ideas. Rather, the intention behind our choice of tracking this singular function is to adduce a classic case of a seemingly isolated function that is thought to be reasonably well understood in terms of peptide structure, before proceeding to an alternative, if not precisely opposite generalized conception of pleiotropic engineering, in which multiple functions are simultaneously optimized into a single peptide sequence. We first examine how membrane activity is encoded into a variety of peptides and proteins: In each of these molecules, membrane interactions underlie their roles in a variety of biological processes that involve membrane remodeling, including antimicrobial activity, membrane fusion and fission, budding and scission, and direct membrane translocation. In an extreme illustrative case of how membrane activity can be combined with other activities, we show how some of the structural requirements for membrane remodeling in innate immunity peptides, such as amphipathicity, can also allow self-assembly *without* membranes into scaffolds that organize immune ligands for multivalent presentation to immune receptors, leading to massive immune amplification. We expand the scope of inquiry and describe how machine-learning-based methods can be used to train a classifier that can discover unanticipated membrane activity in other proteins, especially proteins or peptides that have been annotated to have other primary functions (such as neuropeptides in signaling or molecular motors for force generation). Not only do we see how nature combines membrane activity with other functions, but we can also see specifically how membrane activity comes into being via evolution by combining machine learning and classical evolutionary biology methods. Finally, with these vignettes as a backdrop, we look at some engineering possibilities where we add membrane activity into existing molecules to make synthetic multifunctional molecules. Within the compass of this review, we use techniques that are not usually applied to bioconjugation problems, such as machine learning and synchrotron X-ray diffraction. The hope is that by using some of these techniques, we can learn by example from nature's strategies of multiplexing different functions into the same molecule.

## 2. MULTIFUNCTIONALITY IN INNATE IMMUNITY PROTEINS

**2.1. AMPs.** Antimicrobial peptides (AMPs) constitute a major component of the innate host defense system.<sup>8–13</sup> Collectively, they display broad-spectrum antimicrobial activity and though widely diverse in sequence and structure, they share some common features. Most AMPs are short (<50 amino acids), cationic, and adopt an amphipathic structure that is characterized by segregated polar/cationic and hydrophobic regions.<sup>8–12</sup> They are usually classified based on their secondary structures, with a large subgroup consisting of linear peptides that adopt an amphipathic  $\alpha$ -helix upon contact with a membrane, such as most cathelicidins<sup>14</sup> and magainins.<sup>15</sup> Another subgroup includes AMPs that consist of  $\beta$ -sheet structures that are stabilized by disulfide bonds, such as defensins<sup>16–18</sup> and protegrins.<sup>19</sup> Other peptides form extended structures that are dominated by a few amino acids, like the tryptophan-rich indolicidin<sup>20</sup> and the proline- and arginine-rich PR-39.<sup>21</sup>

In general, AMPs function by disrupting the integrity of bacterial cell membranes, which leads to depolarization, leakage, and cell death.<sup>8,9</sup> Membrane destabilization by AMPs is believed to be a consequence of their cationic and amphipathic nature, which promotes interactions between the peptide and the cell membrane. More specifically, electrostatic interactions between the polar/cationic residues of the AMP and the anionic membrane surface cause peptide–membrane attraction and binding. After adsorption of the AMP onto the membrane, hydrophobic interactions between the hydrophobic residues of the peptide and the bilayer interior perturb packing of the membrane lipids.<sup>8–10,22,23</sup> Together, this combination of interactions leads to membrane disruption that can manifest in a variety of ways, including pore formation,<sup>22,24–26</sup> blebbing,<sup>27,28</sup> budding,<sup>29</sup> and vesicularization.<sup>30</sup> It is important to note that peptide concentrations *in vivo* can be several orders of magnitude greater than those shown to induce membrane disruption *in vitro*.<sup>16,31</sup> Accordingly, these peptides potentially have greater capacity for membrane disruption than what has been demonstrated in experiments. In addition, increasing evidence indicates that certain AMPs utilize alternative modes of action, such as translocating across the cell membrane to interact with intracellular targets and consequently interfering with cellular processes. For example, indolicidin is able to permeabilize membranes without causing cell lysis. Instead, it kills bacteria by binding to DNA and inhibiting DNA synthesis.<sup>32</sup> Although in such cases, membrane permeabilization alone may not achieve bacterial killing, it is still necessary in order for the AMP to reach its cytosolic targets.

While the membrane-disruptive function of AMPs is critical in defending against bacterial infections, more recent studies have also found that the role of AMPs in host defense is not limited to operating as direct microbicides. In fact, many AMPs have been discovered to also function in modulating the immune response, which can occur through activities such as inducing chemotaxis and chemokine production, inhibiting pro-inflammatory cytokine production, and modulating TLR-dependent inflammatory responses.

**2.1.1. Chemotactic Activity.** Some AMPs have exhibited functions outside of membrane activity, and can directly recruit leukocytes or induce the expression of cytokines/chemokines, which can in turn indirectly recruit effector cells, including neutrophils, monocytes, macrophages, immature dendritic cells (DCs), and T cells. For instance, both human  $\alpha$ - and  $\beta$ -defensins are chemotactic for memory T cells and immature DCs, which is believed to occur through interactions with the G-protein-coupled receptor CCR6.<sup>33,34</sup> Human  $\alpha$ -defensins HNP-1 and HNP-2 display chemotactic activity that mediates recruitment of monocytes to sites of inflammation,<sup>35</sup> while human  $\beta$ -defensins HBD-3 and HBD-4 are reported to be chemotactic for monocytes and macrophages,<sup>36</sup> and HBD-2 has chemoattractant activity for mast cells.<sup>37</sup> Cathelicidins have similarly been found to exert chemotactic activity. For humans, LL-37 is chemotactic for neutrophils, monocytes, and T cells. In some cases, its chemotactic activity, such as for human blood-derived monocytes and T cells, is mediated by the G-protein-coupled formyl peptide receptor-like 1 (FPRL-1).<sup>38</sup> Cathelicidins from bovine, human, mouse, and pig are chemotactic for nearly all subsets of peripheral blood cells.<sup>39–41</sup> Granulysin, an AMP that is released by cytotoxic T cells and NK cells, exhibits potent broad-spectrum antimicrobial activity against both Gram-positive and Gram-negative bacteria.<sup>42,43</sup> In addition to its microbicidal activity, granulysin

also has demonstrated chemotactic activity toward monocytes, memory T cells, NK cells, and mature dendritic cells.<sup>44</sup>

AMPs have also been shown to have indirect chemotactic effects by stimulating the production of cytokines and chemokines from a range of cell types via receptor-dependent mechanisms.<sup>33,45</sup> LL-37 induces release of IL-6, IL-8, TNF- $\alpha$ , and granulocyte-macrophage colony-stimulating factor, and IL-1 $\beta$  in human keratinocytes,<sup>46</sup> and enhances the IL-1 $\beta$ -induced secretion of cytokines IL-6, IL-10, and chemokines MCP-1 and MCP-3 in human peripheral blood monocytes.<sup>47</sup> Its induction of IL-8 secretion then promotes the chemotaxis of neutrophils and release of LL-37 at high concentrations.<sup>48</sup> HBD-2, HBD-3, HBD-4, and LL-37 interact with G-protein-coupled receptors and increase expression of IL-6, IL-10, IP-10, MCP-1, MIP-3 $\alpha$ , and RANTES in human keratinocytes.<sup>49</sup> Furthermore, despite their structural differences, both human  $\beta$ -defensins and LL-37 have been found to exhibit similar effects on the activation of the ERK mitogen activated protein kinase (MAPK) pathway to stimulate pro-inflammatory cytokine production in epithelial cells.<sup>49,50</sup> Likewise, granulysin activates monocytes and U937 cells to secrete proinflammatory cytokines, such as TNF- $\alpha$ , MCP-1, MCP-3, MIP-1 $\beta$ , RANTES, IL-1, IL-6, IL-10, and IFN- $\gamma$ .<sup>44</sup>

**2.1.2. Inhibition of Pro-Inflammatory Cytokines.** AMPs can also suppress certain cellular responses to inflammatory stimuli in order to offer protection against potentially lethal effects. For example, LL-37 can inhibit the secretion of pro-inflammatory cytokines induced by bacterial endotoxins, such as lipopolysaccharide (LPS) and lipoteichoic acid (LTA), to protect against endotoxemia.<sup>51–53</sup> It has been further proposed that LL-37 is able to bind and neutralize the endotoxins before they can trigger inflammation.<sup>53–56</sup> Similar protective effects against LPS-induced cytokine production have been observed for defensins and other cationic peptides.<sup>57</sup>

**2.1.3. Modulation of TLR-Dependent Inflammatory Responses.** Toll-like receptors (TLRs) are widely expressed pattern-recognition receptors (PRRs) that detect signature pathogen-associated molecular patterns (PAMPs) from microorganisms and trigger inflammatory responses. These TLR-dependent inflammatory responses have been shown to be modulated by AMPs. Previous work has found that in allergic contact dermatitis, cathelicidins downregulate inflammation by inhibiting TLR4-mediated induction of cytokines in DCs, as well as inhibiting TLR4-induced DC maturation, which includes inhibiting the upregulation of costimulatory molecules CD40, CD80, and CD86.<sup>58</sup> As described above, LL-37 can suppress the endotoxin-stimulated production of pro-inflammatory cytokines. This anti-inflammatory effect has been attributed to the inhibition of LTA-induced TLR2 activation and LPS-induced TLR4 activation.<sup>51,57</sup>

Alternatively, the effects of LL-37 exposure on TLR responses can also be pro-inflammatory. For instance, LL-37 is able to significantly enhance the flagellin activation of TLR5 in keratinocytes,<sup>59</sup> as well as TLR activation by nucleic acids, namely TLR9 by DNA,<sup>60–62</sup> TLR3 by double-stranded RNA (dsRNA),<sup>63,64</sup> and TLR7/TLR8 by single-stranded RNA (ssRNA).<sup>65</sup> In particular, LL-37 has been found to complex with the DNA and RNA ligands via electrostatic interactions, which triggers TLR activation and secretion of pro-inflammatory cytokines. In fact, this phenomenon has been observed to occur across a wide range of molecules, including other AMPs. High-resolution structural studies using small-angle X-ray scattering (SAXS) showed that cationic molecules can form

columnar nanocrystalline complexes with dsDNA and dsRNA that amplify TLR9 and TLR3 activation, respectively, the degree of which is quantitatively correlated with the measured spacing between the nucleic acid columns of the complexes.<sup>64,66,67</sup> Depending on the identity of the cationic molecule, this spacing parameter will vary. However, an optimal spacing was found to exist for both DNA and RNA that resulted in the strongest amplification of TLR-dependent cytokine production. Most strikingly, this spacing approximately matches the steric size of the TLR studied, suggesting that multivalent intercalative ligand–TLR binding is involved in amplifying TLR activation and cytokine production.<sup>68</sup>

**2.2. Immune-Signaling Molecules.** While AMPs in general can exert their antimicrobial effects through direct membrane-disruptive activity, we have highlighted a number of complementary alternative functions of AMPs that can selectively modulate the immune defense responses against bacterial infections. The next question to ask is whether the opposite also applies, in which traditionally recognized immune-signaling molecules function as antimicrobial agents via membrane destabilization. Increasing evidence over recent years indicates that many of such molecules do indeed display direct bactericidal activity, which then suggests that they could have been classified as AMPs if this was their initially identified function. Interestingly, many of these immune molecules feature structural and compositional attributes that are common to AMPs.

For instance, the “kinocidins” constitute a class of chemokines, small secretory cytokines involved in chemotaxis, that have been discovered to possess direct microbicidal activity complementary to their ability to modulate inflammatory responses.<sup>69–73</sup> In fact, many chemokines contain a  $\beta$ -sheet rich “ $\gamma$ -core” motif that also exists in classical AMPs such as defensins,<sup>71,73,74</sup> as well as cationic charge and amphipathicity.<sup>72,74</sup> Moreover, many of these immune signaling molecules have structures that also integrate modular domains of  $\alpha$ -helices and  $\beta$ -sheets, which parallel those of AMPs. Collectively, these similarities point to evolutionary conservation of membrane-disruptive motifs that provide additional functionality and increased efficacy of host defense against pathogens. In addition, the chemotactic activity of certain AMPs further suggests functional reciprocity between chemokines and AMPs. Chemokines are classified into four groups based on conserved N-terminal cysteine motifs: CXC, CC, C, and CX<sub>3</sub>C.<sup>75</sup> Examples of kinocidins include platelet factor 4 (PF-4), also known as platelet chemokine (CXC motif) ligand 4 (CXCL4),<sup>72,76</sup> platelet basic protein (PBP),<sup>77</sup> several IFN- $\gamma$ -inducible CXC chemokines,<sup>78</sup> and interleukin-8 (IL-8), also called chemokine (CXC motif) ligand 8 (CXCL8), one of the most well-characterized and recognized for its role in neutrophil chemotaxis. Despite its similarity to other kinocidins, IL-8 was only recently identified to have direct antimicrobial activity.<sup>71,79</sup> Although most cytokines that have been found to possess direct antimicrobial activity are chemokines, bactericidal interleukins and interferons also exist. For instance, interleukin-26 (IL-26), a human Th17 cell-derived cytokine, was shown to kill bacteria by porating their membranes, while also being able to form complexes with both bacterial and self-DNA to activate TLR9 and induce production of pro-inflammatory cytokines.<sup>80,81</sup> Another study demonstrated that type I interferon- $\beta$  (IFN- $\beta$ ), which is most known for its role in the immune response against viral infections,<sup>82</sup> can also directly kill Gram-positive bacteria, such



as *Staphylococcus aureus*, via membrane permeabilization.<sup>83</sup> This result supports the accumulating evidence that points to the involvement of type I IFNs in host response to bacterial infections,<sup>84–86</sup> along with viral infections. Most striking is that the structures of both IL-26<sup>81</sup> and IFN- $\beta$ <sup>83</sup> contain domains with amphipathic  $\alpha$ -helices rich in cationic residues, which are classic attributes of many membrane-disruptive AMPs, and thus believed to confer antimicrobial activity to the two cytokines. Taken together, these findings reveal a previously unrecognized role for cytokines in host defense against bacterial infections that involves direct membrane activity, distinct from their established immune signaling functions.

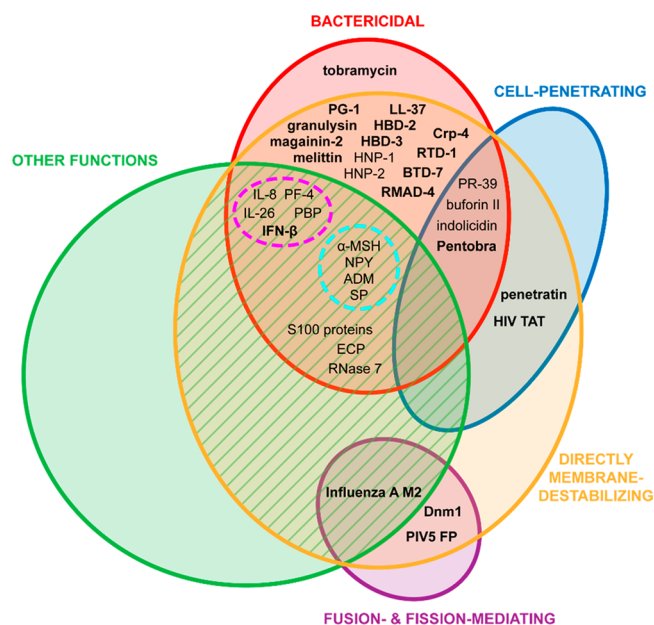
### 3. DIVERSITY OF MEMBRANE-ACTIVE MULTIFUNCTIONAL PROTEINS

The selected examples above illustrate the involvement of immunity molecules in multiple biological functions. In general, one of these is often direct membrane disruption that ultimately results in antimicrobial effects. However, protein multifunctionality that encompasses such direct membrane activity is not limited to immune proteins, and certain proteins may also exert membrane-disruptive and immunomodulatory effects in addition to their other known functions (Figure 1).

**3.1. S100 Family.** The S100 family of EF-hand calcium-binding proteins regulate a wide range of intracellular and extracellular functions, including proliferation, differentiation, apoptosis, metabolism, and autocrine and paracrine signaling.<sup>87</sup> Collectively, they have been discovered to exert alternative functionality as immunomodulators and direct microbicides. Several members of the family, namely S100A8, S100A9, and S100A12, have been reported to produce pro-inflammatory effects and are implicated in a variety of inflammatory conditions, such as psoriasis, arthritis, and chronic inflammatory bowel disease.<sup>88–90</sup> Studies have specifically found that both S100A8<sup>91</sup> and S100A12<sup>92</sup> can modulate host immune responses through TLR4 activation, and that S100A12 also exhibits chemotactic activity for monocytes.<sup>93</sup> Furthermore, members of the S100 family are characterized by helix-rich structures and have been described to interact with lipids. With structural similarity to membrane-active antimicrobial molecules, it is not surprising that S100A7<sup>94</sup> and S100A12<sup>92</sup> have the capacity to kill bacteria through membrane permeabilization.

**3.2. RNase A Superfamily.** The ribonuclease (RNase) A superfamily has been intensively studied and consists of vertebrate RNase homologues to bovine pancreatic ribonuclease (RNase A).<sup>95</sup> Despite a wealth of information on the structural and chemical properties of RNase A family members, most of their biological functions remain unclear. In addition, members of the RNase A superfamily exhibit considerable sequence divergence, with identities that range from 20% to nearly 100%.<sup>96</sup> In humans, eight RNases have been identified, including several that are antimicrobial.<sup>96</sup> Among these, RNase 3 and RNase 7 are two representative members with well-characterized bactericidal activity against a variety of pathogens. These antimicrobial RNases are small, highly cationic, and share the common RNase A structure and key residues required for their enzymatic function. However, evidence indicates that their bactericidal activity is associated with membrane disruption and do not depend on their conserved catalytic RNase activity.<sup>96–99</sup>

Eosinophil cationic protein (ECP), a member of the RNase A superfamily and also known as RNase 3, is one of the major



**Figure 1.** Diagram illustrating diversity among membrane-active proteins and peptides. While bactericidal (red region), cell-penetrating (blue region), and membrane fusion- and fission-mediating (violet region) proteins and peptides are highly diverse, their functions often involve the ability to directly destabilize membranes (gold region). Of particular interest are membrane-active multifunctional proteins (green hashed region), which are those generally recognized for having other main functions (green region) in addition to roles associated with membrane-destabilizing activity. Thus far, the relatively low number of proteins identified as multifunctional has been attributed to the serendipitous nature of their discovery. However, newly developed machine-learning tools allow for systematic large-scale exploration of new and existing protein taxonomies to uncover additional membrane-active multifunctional proteins. Proteins and peptides in bold have been shown to generate negative Gaussian curvature (NGC) using SAXS experiments. Specific cytokines (dashed magenta circle) and neuropeptides (dashed cyan circle) that have been identified to be antimicrobial are also indicated. Table S1 provides references for the proteins and peptides shown here.

granule proteins secreted from activated eosinophils. Eosinophils have been implicated in bacterial infections and inflammation, as well as in immunoregulation and tissue remodeling.<sup>100,101</sup> Remarkably, ECP is a single-chain cationic polypeptide that displays potent, broad-spectrum bactericidal activity against both Gram-positive and Gram-negative strains, which is believed to occur through a mechanism of action involving membrane disruption and pore formation.<sup>102–104</sup> ECP has specifically been shown to permeabilize the membranes of *Escherichia coli*, *S. aureus*, and *Mycobacterium vaccae* and create pores in lipid vesicles.<sup>97,102–106</sup> Recent studies have attributed its antimicrobial activity to an N-terminal domain that bears striking structural and chemical similarity to AMPs, such as adopting an  $\alpha$ -helical conformation and having a net positive charge due to a high number of arginine residues.<sup>105</sup> In fact, both hydrophobic and cationic residues have been found to play key roles in the membrane-disruptive bactericidal activity of ECP by promoting protein binding and insertion into lipid bilayers.<sup>104,107</sup> Aside from exerting antimicrobial effects, ECP has also been reported to participate in immunomodulation and inflammation in asthma and allergic diseases.<sup>108,109</sup>

RNase 7 is expressed in epithelial tissues and has similarly been described to exhibit both broad-spectrum antimicrobial activity and RNase activity. Along with other skin-derived AMPs, RNase 7 is considered one of the main components of the host defense against cutaneous infections.<sup>96,110</sup> Notably, RNase 7 has been shown to be effective against both Gram-positive (*S. aureus*, *Staphylococcus saprophyticus*, *Propionibacterium acnes*, *Enterococcus faecalis*, vancomycin-resistant *Enterococcus faecium*) and Gram-negative bacteria (*E. coli*, *Pseudomonas aeruginosa*, *Klebsiella pneumoniae*, *Proteus mirabilis*).<sup>110,111</sup> Like ECP, the bactericidal activity of RNase 7 has been associated with its capacity to bind and permeate membranes, with surface clusters of positively charged lysine residues playing a crucial role.<sup>98</sup> Interestingly, the exposure of keratinocytes to bacteria induces RNase 7 expression, a response that has been observed for other epithelial AMPs such as HBD-2, HBD-3, and HBD-4.<sup>110</sup>

These findings together suggest that ECP and RNase 7 share a common bactericidal mechanism of action involving membrane disruption. While both proteins are cationic with an abundance of arginine and lysine surface-exposed residues and display broad antimicrobial activity against Gram-positive and Gram-negative bacteria, they only share 40% sequence identity. Indeed, members of the RNase A superfamily exhibit extensive diversity, despite all having conserved catalytic RNase activity. The antimicrobial properties of ECP and RNase 7 therefore not only support the role of RNase A family members in host defense, but also underscore the multifunctional diversity that exists within the superfamily.

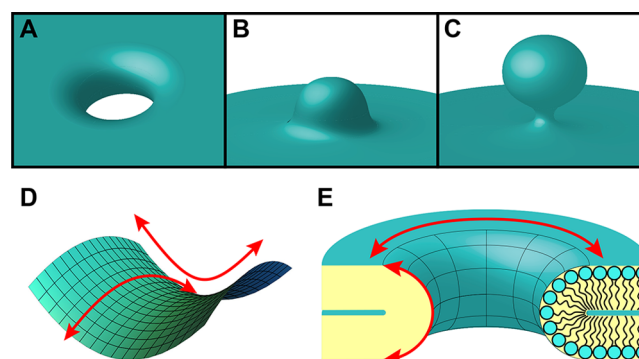
#### 4. GENERAL PHYSICOCHEMICAL AND STRUCTURAL PROPERTIES OF MEMBRANE-ACTIVE PEPTIDES

While the antimicrobial activity of AMPs via membrane destabilization has been ascribed to their shared fundamental amphipathic motif, a large body of work has further examined how specific physicochemical properties and amino acid sequence composition can contribute to their bactericidal activity. In general, these factors influence the electrostatic and hydrophobic peptide–membrane interactions that overall determine AMP activity.

Among  $\alpha$ -helical AMPs, for example, which constitute the largest and most extensively studied group of AMPs, those with higher cationic charge typically have greater affinity to more negatively charged bacterial membranes and therefore tend to be more selective for bacterial over mammalian cells.<sup>9,112</sup> Accordingly, increased cationic charge, to a certain degree, has been demonstrated to enhance both antimicrobial activity and selectivity.<sup>113–115</sup> Studies have also similarly shown that increased hydrophobicity can increase bactericidal activity, however, at the expense of selectivity.<sup>116,117</sup> These findings suggest that an optimal balance of charge and hydrophobicity exists for maximal antimicrobial efficacy. Not only is the overall hydrophobicity important for antimicrobial activity, but additional properties such as the hydrophobic moment, often described as amphipathicity, and the angle subtended by hydrophobic helix face also correlate with bactericidal activity.<sup>118,119</sup> For instance, helical AMPs generally feature a wide hydrophobic face and a narrow polar face, which allow them to penetrate into the membrane interior and disrupt lipid packing.<sup>120</sup> In fact, increasing the bulkiness or the angle subtended by the hydrophobic face enhances the ability of the AMP to disrupt membranes and induce lysis.<sup>113,118,121</sup>

Further investigation into the relationship between charge and hydrophobicity included a compositional analysis of over 1080 cationic helical AMPs from the antimicrobial database.<sup>122</sup> This study identified a positive correlation between  $N_K/(N_K + N_R)$  (the ratio of the number of lysines to total number of lysines and arginines) and the average peptide hydrophobicity based on the Eisenberg consensus scale.<sup>112,123</sup> Because membrane disruption is the general mechanism for AMPs, this relationship is informative about the relative amounts of arginine, lysine, and hydrophobicity that are needed on average to confer membrane activity to a helical peptide. We point out that a corollary of this argument is that membrane activity underdetermines the full sequence of an AMP, so that additional functions can be encoded into the same sequence.

Although the actual process of membrane disruption by an AMP depends on the specific peptide and target membrane, AMPs have been found to destabilize membranes through a range of modes such as pore formation,<sup>22,24–26</sup> blebbing,<sup>27,28</sup> budding,<sup>29</sup> and vesicularization.<sup>30</sup> Interestingly, the generation of saddle-splay membrane curvature, also known as negative Gaussian curvature (NGC), is topologically required for all of these membrane-destabilizing processes (Figure 2).<sup>124</sup> This



**Figure 2.** Negative Gaussian curvature (NGC). NGC manifests in a variety of membrane-remodeling processes, including (A) along the interior of a transmembrane pore, (B) at the bases of blebs, and (C) at the necks of buds. (D) NGC is also known as saddle-splay curvature due to the shape of its surface, which bends upward in one direction and downward in the orthogonal direction (red arrows). (E) A cross-section of a transmembrane pore depicts its curved inner surface that is characterized by NGC. The directions of the curvatures along the membrane surface that together create NGC are represented by red arrows.

type of membrane curvature is characterized by a surface that bends upward in one direction and bends downward in the orthogonal direction. Recent work using SAXS measurements reported that the ability of AMPs, both  $\alpha$ -helical and  $\beta$ -sheet peptides, to disrupt and permeabilize bacterial membranes is correlated with their capacity to induce NGC in lipid bilayers.<sup>112,123,125</sup> The generation of NGC by AMPs is believed to arise from the combination of distinct membrane curvature effects produced by the electrostatic and hydrophobic peptide–membrane interactions.<sup>112,123</sup> While the ability to generate NGC has also been observed for AMP mutants<sup>112,125</sup> and synthetic AMP mimics,<sup>126–129</sup> it is not limited to antimicrobial agents. For many other molecules with functions that involve membrane destabilization, a strong correlation has also been found between NGC generation and their activity. For instance, the induction of NGC is also exhibited by a range of cell-penetrating peptides (CPPs), a cognate class of molecular pore

formers.<sup>130–132</sup> CPPs are short peptide sequences capable of efficient translocation across cell membranes, while remaining nonlytic.<sup>133,134</sup> Due to this unique ability, CPPs are routinely utilized for mediating the intracellular delivery of attached macromolecules. Most CPPs are cationic, with the arginine-rich CPPs, such as the peptide derived from the human immunodeficiency virus (HIV-1) transactivator of transcription (TAT) protein,<sup>131,135,136</sup> being the most widely studied. Some CPPs can also adopt amphipathic structures, as seen with penetratin.<sup>132,137</sup> While their exact mechanism remains controversial, evidence indicates that cellular uptake of CPPs can involve more than one pathway,<sup>138–140</sup> such as direct translocation<sup>141–143</sup> and endocytosis.<sup>144,145</sup> Nonetheless, for any of these modes of action, membrane destabilization must occur for CPPs to enter cells. Consistent with the trend identified for AMPs, CPPs can also generate NGC in lipid membranes.<sup>130–132,146</sup> In addition to AMPs and CPPs, other peptides directly involved in membrane-remodeling processes have also been shown to induce this specific type of destabilizing membrane curvature, including viral budding peptides<sup>147</sup> and viral fusion peptides.<sup>148</sup>

## 5. DISCOVERY OF COMPLEMENTARY MEMBRANE ACTIVITY IN DIVERSE PROTEINS AND PEPTIDES

The ability to generate NGC has been observed for a diverse range of proteins and peptides associated with membrane-remodeling activity, which collectively suggests that the generation of NGC is a common root mechanism for membrane-destabilizing processes.

Furthermore, the detection of NGC-generating ability in other proteins and peptides can point to the existence of complementary membrane-destabilizing activity that has not been previously recognized or associated with their established function. Indeed, some proteins are now described as multifunctional due to recent discoveries of their additional antimicrobial activity. IFN- $\beta$ , a cytokine known for its role in antiviral defense,<sup>82</sup> was found to be directly antimicrobial against Gram-positive bacteria.<sup>83</sup> Experiments demonstrated that the microbicidal activity of IFN- $\beta$  stems from its ability to bind and permeabilize bacterial membranes. Remarkably, an  $\alpha$ -helical domain of the cytokine closely resembles classical membrane-disruptive AMPs with respect to amino acid composition, amphipathicity, cationic charge, hydrophobicity, and the ability to generate NGC in membranes. S100A12 is another example in which AMP-like sequences exhibit bactericidal activity. As a member of the S100 family of calcium-binding proteins, this host-defense protein is associated with the inflammatory response and mediates a variety of cellular processes.<sup>87</sup> S100A12 has been reported to display antimicrobial activity against bacteria.<sup>92,149</sup> Interestingly, the structure of S100A12 contains multiple  $\alpha$ -helices characterized by cationic charge and amphipathicity. Compositional analysis further revealed that the amino acid content of S100A12 follows the sequence trend of known membrane-disruptive helical AMPs and fulfills the criterion for NGC generation.<sup>92,123</sup> The newly discovered antimicrobial activities of IFN- $\beta$  and S100A12 are only a sample of the functional diversity that exists within the realm of immunity proteins, and together suggest that far more proteins with multiplexed functionalities remain unidentified.

As we have observed, the membrane-disruptive bactericidal activity of AMPs depends on a variety of factors, many of which are closely related and may be interdependent.<sup>150</sup> Due to this

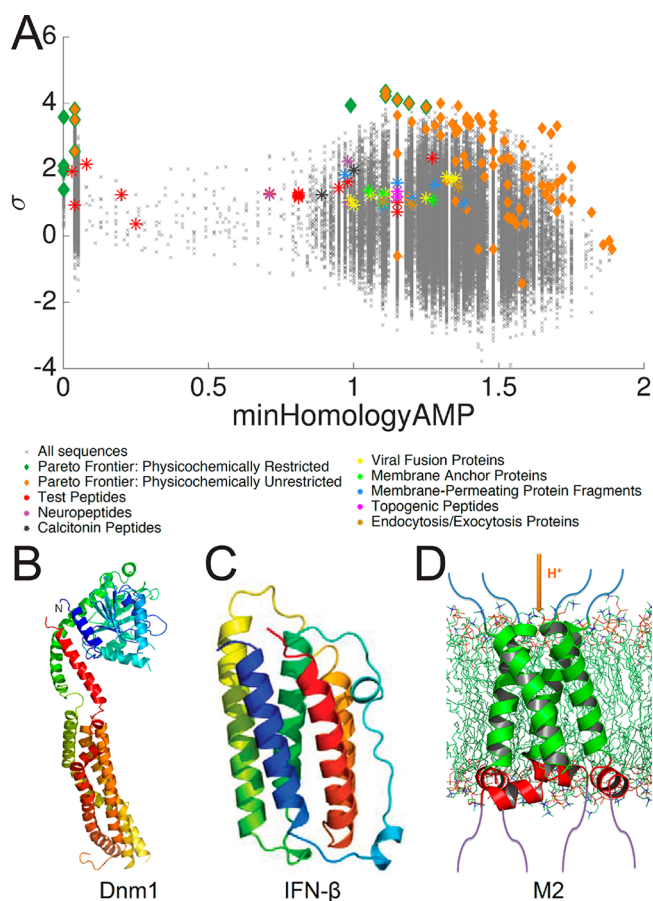
complexity, simple correlations between AMP activity and individual peptide properties are unlikely. In light of this, we expanded upon our earlier work, which identified correlations between amino acid composition and hydrophobicity in membrane-disruptive AMPs as well as between membrane-destabilizing effects and NGC generation, to develop a machine-learning algorithm that effectively predicts the membrane-destabilizing and NGC-generating capacities of a peptide based on its sequence and associated physicochemical properties.

## 6. DISCOVERING MEMBRANE-ACTIVE PEPTIDES AND PROTEINS USING MACHINE LEARNING

Given the sequence and structural diversity of membrane-active peptides and proteins, it is difficult to detect hidden membrane activity without direct empirical testing, apart from homology searches and sequence/structural alignments. However, the emergence of quantitative structure–activity relationship (QSAR) computational tools has enabled large scale in silico screening of genomes for membrane-active peptides and proteins. For example, many QSAR methods have been applied to discover and design membrane-active AMPs, a prototypical class of membrane-active peptides.<sup>151–155</sup> Recently, a support vector machine (SVM)-based machine-learning tool was developed to identify peptide sequences that generate NGC in membranes.<sup>156–158</sup> The SVM was trained on a database of experimentally validated  $\alpha$ -helical AMPs<sup>122,159,160</sup> and decoy non-AMP  $\alpha$ -helical peptides.<sup>161</sup> Based on 12 physicochemical descriptors most predictive of antimicrobial activity, the machine-learning tool takes a peptide sequence as input, and outputs a prediction of antimicrobial activity. Additional calibrating SAXS experiments on model membranes identified a relationship between the machine-learning classification metric ( $\sigma$ ) and the ability of a peptide sequence to induce negative Gaussian membrane curvature.<sup>156</sup> This machine-learning tool not only can guide the design of new synthetic membrane-active peptides for therapeutic purposes, but also allows us to probe the sequence space for previously unrecognized underlying membrane activity in new and existing protein taxonomies (Figure 3). In fact, the machine-learning classifier has successfully predicted membrane activity in neuropeptides, viral fusion proteins, endocytosis/exocytosis machinery, and mitochondrial fission proteins, among many others.<sup>156</sup> Because of the serendipitous nature of multifunctional protein discovery, the number of proteins that have been identified and characterized as such is not high. However, this newly developed machine-learning tool now enables a systematic large-scale in silico approach for discovering protein sequences with membrane activity.

The machine-learning classifier has correctly predicted membrane-destabilizing activity in sequences contained within proteins that have been generally associated with other activities. These include cytokine IFN- $\beta$ ,<sup>83</sup> mitochondrial fission protein Dnm1,<sup>162</sup> and the influenza A virus M2 protein.<sup>147</sup> IFN- $\beta$  consists of a six-helix bundle tertiary structure stabilized by cysteines. Electrostatic surface potential calculations revealed that several component helices of IFN- $\beta$  are cationic and amphipathic, similar to canonical helical AMPs.<sup>83</sup> Using a moving window scan, the component helices of IFN- $\beta$  were screened for membrane activity using the machine-learning classifier, which predicted helix 4 of IFN- $\beta$  to possess antimicrobial activity with high probabilities ( $\sigma = 0.95$ ,  $P(+1) = 0.960$ ). This finding is consistent with SAXS measurements that





**Figure 3.** Machine learning discovers hidden NGC-generating activity in proteins with diverse primary functions. (A) A machine-learning-based Monte Carlo search of the undiscovered peptide sequence space reveals sequences in existing peptides and proteins with the ability to remodel membranes.<sup>156</sup> (B) Dnm1 is a molecular motor GTPase that also generates NGC to facilitate mitochondrial fission.<sup>162</sup> (C) IFN- $\beta$  is a pleiotropic, immunomodulatory cytokine that exhibits direct antimicrobial activity and NGC generation.<sup>83</sup> (D) Influenza A M2 protein is a proton-selective ion channel that also generates NGC to enable viral budding and scission.<sup>147</sup> All figure panels are reproduced with permission from their respective sources.

showed the ability of the helix to induce NGC in model membranes and antimicrobial assays that demonstrated its membrane-disruptive microbicidal activity against Gram-positive bacteria.<sup>83</sup>

Dnm1 is a cytosolic dynamin-related GTPase that plays a key role in the regulation of yeast mitochondrial fission.<sup>163</sup> The current model describes Dnm1 purely as a molecular motor, forming spirals that wrap around mitochondria and constrict upon GTP hydrolysis to ultimately cause pinching and scission of the mitochondrial membrane. Using the machine-learning classifier, an N-terminal helix of Dnm1 was predicted to have membrane-destabilizing activity.<sup>162</sup> SAXS experiments also found this specific helix to have the capacity to generate NGC in model mitochondrial membranes and promote the formation of fission necks narrower than could be achieved with simply mechanical pinching.<sup>162</sup> These results together suggest that Dnm1 catalyzes mitochondrial fission through the combination of mechanical constriction and synergistic membrane-remodeling activity. Machine-learning analysis of 33 phylogenetically clustered Dnm1 relatives further revealed that many members within the dynamin superfamily had

evolved the capacity to generate NGC and destabilize membranes.

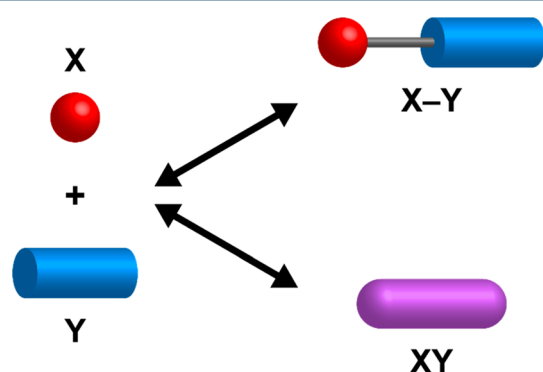
Recent efforts have identified an additional function of the influenza A virus M2 protein in directly inducing membrane curvature that promotes viral budding activity. During the viral replication cycle, viral components assemble to form progeny virions that bud at the plasma membrane of the infected host cell, thereby spreading infection. The budding of enveloped viruses is a complex process that involves the extrusion of the membrane and scission of the bud to release the virion from the former host, which are steps that require organized membrane remodeling with localized regions of high membrane curvature. While many enveloped viruses, such as HIV-1 and the Ebola virus, hijack the host endosomal sorting complex required for transport (ESCRT) machinery to facilitate membrane scission and budding virion release,<sup>164–166</sup> studies have suggested that the influenza virus is able to undergo budding via an ESCRT-independent mechanism.<sup>165,167</sup> The M2 protein from the influenza A virus is a multifunctional protein that forms a homotetramer in the membrane to function as a proton-selective ion channel<sup>168,169</sup> and plays a variety of roles during the life cycle of the virus.<sup>170–172</sup> For instance, *in vitro* and *in vivo* experiments have revealed that the M2 protein mediates budding and virion release from cells.<sup>173,174</sup> Interestingly, the predominant localization of M2 at the necks of budding virions points to its ability to directly induce the specific types of membrane curvature that facilitate budding and membrane scission.<sup>173,175</sup> Indeed, results from SAXS measurements of M2-induced membrane deformations showed that the protein generates NGC in model membranes.<sup>147</sup> Consistent with its role in promoting the final pinching off of a budding virion, the quantitative amount of NGC induced by M2 can produce a scission neck with a diameter that is ten times smaller than the diameter of a spherical budding virion.<sup>147</sup> Remarkably, the generation of NGC by M2 is largely conferred by its cytoplasmic C-terminal amphipathic helix, which has been shown using electron microscopy,<sup>173</sup> SAXS measurements,<sup>147</sup> and machine-learning classifier<sup>156</sup> predictions ( $\sigma = 1.63$ ,  $P(+1) = 0.996$ ). More specifically, the C-terminal amphipathic helical domain of M2 alone is sufficient to generate NGC, however, to a lesser extent than the full-length protein.<sup>147</sup> This finding indicates that other regions of the M2 protein affect curvature generation, which further suggests that cross-talk between different domains within multifunctional proteins is likely involved in achieving maximal protein activity.

The examples presented here demonstrate that the machine-learning classifier is a powerful tool that can efficiently and accurately detect membrane activity, and therefore, can enable the discovery of membrane-active multifunctional proteins that have so far been primarily recognized for other functions.

## 7. DESIGN OF SYNTHETIC MULTIFUNCTIONAL MOLECULES

The ability to design in membrane-disruptive activity allows for the creation of new multifunctional molecules and provides many opportunities for future therapeutics. It is important to note that the potential influence of cross-talk between domains on the activity of a whole protein presents significant implications for engineering multifunctional proteins, peptides, and their derivatives. On these grounds, we look to nature for guidance. What can we learn from existing multifunctional proteins as products of natural bioconjugation? It would make sense that the most efficient and optimized synthetic designs

would be those that best emulate nature's approach to incorporating multifunctionality into proteins: maximizing usage of the whole molecule to integrate each individual functionality, rather than simply joining multiple modular domains that each have a discrete function (Figure 4).



**Figure 4.** Design of multifunctional molecules. A simple approach to creating a multifunctional molecule is through joining two (or more) components that each have a separate function. For example, component X, depicted here as a red sphere, can be directly conjugated to a second component Y, represented by a blue cylinder, to form the multifunctional compound X–Y (upper right) with modular domains. Alternatively, hybridizing the two components X and Y to create one compound XY (violet spherocylinder, lower right) that integrates the dual functionalities of X and Y within the same domain is a strategy that more closely mimics the design of natural multifunctional molecules.

Taking these factors into consideration, we aimed to create a multifunctional molecule by hybridizing two separate compounds with different functions. More specifically, this strategy was applied in re-engineering conventional antibiotics to target bacteria that have low metabolic activity, known as persisters, which are often tolerant to common antibiotic treatments and play a role in antibiotic resistance.<sup>176–181</sup> Aminoglycoside antibiotics, such as tobramycin, target bacterial ribosomes to block protein synthesis. This class of antibiotics has limited effectiveness against persisters, which have diminished membrane permeability due in part to decreased uptake activity in their low metabolic state. Reduced uptake activity limits the cellular accumulation of drug and consequently decreases susceptibility of the cell to the antibiotic, in turn promoting the development of bacterial resistance to aminoglycosides. Augmenting the membrane permeability of aminoglycosides provides a means to improve their efficacy against persister cells and resistant strains of bacteria. This can be accomplished by conjugating the antibiotic with a short peptide transporter sequence that promotes rapid membrane penetration. The specific design of the transporter sequence is informed by previously established sequence principles for peptides that disrupt and permeate membranes via destabilizing curvature generation.<sup>112,123,131,132</sup> For example, the conjugation of a 12-amino-acid peptide sequence derived from the CPP penetratin<sup>182</sup> with the aminoglycoside tobramycin formed the antibiotic–peptide hybrid Pentobra.<sup>179–181</sup> The compositional contributions from both tobramycin and peptide components of Pentobra together achieve the appropriate proportions of cationic and hydrophobic groups to effectively generate NGC and penetrate cell membranes.<sup>179</sup> By enabling the efficient cellular delivery of aminoglycoside tobramycin, which alone can have limited cell uptake, this hybrid molecule thus displays

multifunctionality by integrating two mutually amplifying bactericidal functionalities: disruption of membrane integrity and inhibition of protein synthesis.

## 8. CONCLUSIONS AND OUTLOOK

In principle, antimicrobial/membrane activity can be grafted onto existing proteins with other functions in a number of ways. The first order solution is to simply conjugate a known membrane-active peptide onto another domain with a different primary function using a covalent linker with conventional bioconjugate chemistry. However, we believe this process can be improved by combining a machine-learning approach with directed evolution or computational protein design methodologies, such as Rosetta. A recent paper utilized a computational method to combine cell-penetrating, DNA-binding, pheromone, and antimicrobial activities into one domain.<sup>183</sup>

The growing number and diversity of identified proteins with multifunctionality suggests that it is a general phenomenon that exists in all kingdoms of life. Expanding our knowledge of protein multifunctionality and its evolution may potentially allow for the selective inhibition or introduction of distinct functions, and thus presents new opportunities for future molecular engineering. The examples shown here illustrate that methods based on machine learning and artificial intelligence can be helpful for informing our understanding of how diverse functions are multiplexed into a single sequence, with implications for the deterministic design of multifunctional molecules.

### ■ ASSOCIATED CONTENT

#### 📄 Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.bioconjchem.8b00176.

Supplementary table of molecules and references (PDF)

### ■ AUTHOR INFORMATION

#### Corresponding Author

\*E-mail: gclwong@seas.ucla.edu.

#### ORCID

Michelle W. Lee: 0000-0003-1613-9501

Ernest Y. Lee: 0000-0001-5144-2552

#### Notes

The authors declare no competing financial interest.

### ■ ACKNOWLEDGMENTS

M.W.L. and G.C.L.W. acknowledge support from NIH Grant 1R21AI122212. E.Y.L. acknowledges support from the Systems and Integrative Biology Training Program (T32GM008185), the Medical Scientist Training Program (T32GM008042), and the Dermatology Scientist Training Program (T32AR071307) at UCLA. E.Y.L. and G.C.L.W. also acknowledge an Early Career Research Grant and a Discovery Grant from the National Psoriasis Foundation, respectively. X-ray research was conducted at Stanford Synchrotron Radiation Lightsource, SLAC National Laboratory, supported by the US DOE Office of Basic Energy Sciences under Contract no. DE-AC02-76SF00515.



## REFERENCES

- (1) Francis, M. B., and Carrico, I. S. (2010) New frontiers in protein bioconjugation. *Curr. Opin. Chem. Biol.* 14, 771–773.
- (2) Stephanopoulos, N., and Francis, M. B. (2011) Choosing an effective protein bioconjugation strategy. *Nat. Chem. Biol.* 7, 876–884.
- (3) Jeffery, C. J. (2016) Protein species and moonlighting proteins: Very small changes in a protein's covalent structure can change its biochemical function. *J. Proteomics* 134, 19–24.
- (4) Apic, G., Gough, J., and Teichmann, S. A. (2001) Domain combinations in archaeal, eubacterial and eukaryotic proteomes. *J. Mol. Biol.* 310, 311–325.
- (5) Jeffery, C. J. (1999) Moonlighting proteins. *Trends Biochem. Sci.* 24, 8–11.
- (6) Jeffery, C. J. (2003) Multifunctional proteins: examples of gene sharing. *Ann. Med.* 35, 28–35.
- (7) Huberts, D. H. E. W., and van der Klei, I. J. (2010) Moonlighting proteins: An intriguing mode of multitasking. *Biochim. Biophys. Acta, Mol. Cell Res.* 1803, 520–525.
- (8) Zasloff, M. (2002) Antimicrobial peptides of multicellular organisms. *Nature* 415, 389–395.
- (9) Shai, Y. (1999) Mechanism of the binding, insertion and destabilization of phospholipid bilayer membranes by  $\alpha$ -helical antimicrobial and cell non-selective membrane-lytic peptides. *Biochim. Biophys. Acta, Biomembr.* 1462, 55–70.
- (10) Brogden, K. A. (2005) Antimicrobial peptides: pore formers or metabolic inhibitors in bacteria? *Nat. Rev. Microbiol.* 3, 238–250.
- (11) Hancock, R. E. W., and Lehrer, R. (1998) Cationic peptides: a new source of antibiotics. *Trends Biotechnol.* 16, 82–88.
- (12) Hancock, R. E. W., and Sahl, H.-G. (2006) Antimicrobial and host-defense peptides as new anti-infective therapeutic strategies. *Nat. Biotechnol.* 24, 1551–1557.
- (13) Yeaman, M. R., and Yount, N. Y. (2003) Mechanisms of Antimicrobial Peptide Action and Resistance. *Pharmacol. Rev.* 55, 27–55.
- (14) Dürr, U. H. N., Sudheendra, U. S., and Ramamoorthy, A. (2006) LL-37, the only human member of the cathelicidin family of antimicrobial peptides. *Biochim. Biophys. Acta, Biomembr.* 1758, 1408–1425.
- (15) Zasloff, M. (1987) Magainins, a class of antimicrobial peptides from *Xenopus* skin: isolation, characterization of two active forms, and partial cDNA sequence of a precursor. *Proc. Natl. Acad. Sci. U. S. A.* 84, 5449–5453.
- (16) Ganz, T. (2003) Defensins: antimicrobial peptides of innate immunity. *Nat. Rev. Immunol.* 3, 710–720.
- (17) Lehrer, R. I. (2004) Primate defensins. *Nat. Rev. Microbiol.* 2, 727–738.
- (18) Selsted, M. E., and Ouellette, A. J. (2005) Mammalian defensins in the antimicrobial immune response. *Nat. Immunol.* 6, 551–557.
- (19) Kokryakov, V. N., Harwig, S. S. L., Panyutich, E. A., Shevchenko, A. A., Aleshina, G. M., Shamova, O. V., Korneva, H. A., and Lehrer, R. I. (1993) Protegrins: leukocyte antimicrobial peptides that combine features of corticostatic defensins and tachyplesins. *FEBS Lett.* 327, 231–236.
- (20) Selsted, M. E., Novotny, M. J., Morris, W. L., Tang, Y. Q., Smith, W., and Cullor, J. S. (1992) Indolicidin, a novel bactericidal tridecapeptide amide from neutrophils. *J. Biol. Chem.* 267, 4292–4295.
- (21) Agerberth, B., Lee, J.-Y., Bergman, T., Carlquist, M., Boman, H. G., Mutt, V., and Jörnvall, H. (1991) Amino acid sequence of PR-39. *Eur. J. Biochem.* 202, 849–854.
- (22) Matsuzaki, K., Sugishita, K.-i., Ishibe, N., Ueha, M., Nakata, S., Miyajima, K., and Epand, R. M. (1998) Relationship of Membrane Curvature to the Formation of Pores by Magainin 2. *Biochemistry* 37, 11856–11863.
- (23) Matsuzaki, K. (1999) Why and how are peptide–lipid interactions utilized for self-defense? Magainins and tachyplesins as archetypes. *Biochim. Biophys. Acta, Biomembr.* 1462, 1–10.
- (24) Spaar, A., Münster, C., and Salditt, T. (2004) Conformation of Peptides in Lipid Membranes Studied by X-Ray Grazing Incidence Scattering. *Biophys. J.* 87, 396–407.
- (25) Yang, L., Harroun, T. A., Weiss, T. M., Ding, L., and Huang, H. W. (2001) Barrel-Stave Model or Toroidal Model? A Case Study on Melittin Pores. *Biophys. J.* 81, 1475–1485.
- (26) Tang, M., Waring, A. J., and Hong, M. (2007) Phosphate-Mediated Arginine Insertion into Lipid Membranes and Pore Formation by a Cationic Membrane Peptide from Solid-State NMR. *J. Am. Chem. Soc.* 129, 11438–11446.
- (27) Saiman, L., Tabibi, S., Starner, T. D., San Gabriel, P., Winokur, P. L., Jia, H. P., McCray, P. B., and Tack, B. F. (2001) Cathelicidin Peptides Inhibit Multiply Antibiotic-Resistant Pathogens from Patients with Cystic Fibrosis. *Antimicrob. Agents Chemother.* 45, 2838–2844.
- (28) Kalfa, V. C., Jia, H. P., Kunkle, R. A., McCray, P. B., Tack, B. F., and Brogden, K. A. (2001) Congeners of SMAP29 Kill Ovine Pathogens and Induce Ultrastructural Damage in Bacterial Cells. *Antimicrob. Agents Chemother.* 45, 3256–3261.
- (29) Yu, Y., Vroman, J. A., Bae, S. C., and Granick, S. (2010) Vesicle Budding Induced by a Pore-Forming Peptide. *J. Am. Chem. Soc.* 132, 195–201.
- (30) Falagas, M. E., Kasiakou, S. K., and Saravolatz, L. D. (2005) Colistin: The Revival of Polymyxins for the Management of Multidrug-Resistant Gram-Negative Bacterial Infections. *Clin. Infect. Dis.* 40, 1333–1341.
- (31) Ayabe, T., Satchell, D. P., Wilson, C. L., Parks, W. C., Selsted, M. E., and Ouellette, A. J. (2000) Secretion of microbicidal  $\alpha$ -defensins by intestinal Paneth cells in response to bacteria. *Nat. Immunol.* 1, 113–118.
- (32) Hsu, C.-H., Chen, C., Jou, M.-L., Lee, A. Y.-L., Lin, Y.-C., Yu, Y.-P., Huang, W.-T., and Wu, S.-H. (2005) Structural and DNA-binding studies on the bovine antimicrobial peptide, indolicidin: evidence for multiple conformations involved in binding to membranes and DNA. *Nucleic Acids Res.* 33, 4053–4064.
- (33) Yang, D., Chertov, O., and Oppenheim, J. J. (2001) Participation of mammalian defensins and cathelicidins in antimicrobial immunity: receptors and activities of human defensins and cathelicidin (LL-37). *J. Leukocyte Biol.* 69, 691–697.
- (34) Yang, D., Chertov, O., Bykovskaia, S. N., Chen, Q., Buffo, M. J., Shogan, J., Anderson, M., Schröder, J. M., Wang, J. M., Howard, O. M. Z., et al. (1999)  $\beta$ -Defensins: Linking Innate and Adaptive Immunity Through Dendritic and T Cell CCR6. *Science* 286, 525–528.
- (35) Territo, M. C., Ganz, T., Selsted, M. E., and Lehrer, R. (1989) Monocyte-chemotactic activity of defensins from human neutrophils. *J. Clin. Invest.* 84, 2017–2020.
- (36) Yang, D., Biragyn, A., Kwak, L. W., and Oppenheim, J. J. (2002) Mammalian defensins in immunity: more than just microbicidal. *Trends Immunol.* 23, 291–296.
- (37) Niyonsaba, F., Iwabuchi, K., Matsuda, H., Ogawa, H., and Nagaoka, I. (2002) Epithelial cell-derived human  $\beta$ -defensin-2 acts as a chemotaxin for mast cells through a pertussis toxin-sensitive and phospholipase C-dependent pathway. *Int. Immunol.* 14, 421–426.
- (38) Yang, D., Chen, Q., Schmidt, A. P., Anderson, G. M., Wang, J. M., Wooters, J., Oppenheim, J. J., and Chertov, O. (2000) LL-37, the Neutrophil Granule- And Epithelial Cell-Derived Cathelicidin, Utilizes Formyl Peptide Receptor-Like 1 (Fpr1) as a Receptor to Chemo-attract Human Peripheral Blood Neutrophils, Monocytes, and T Cells. *J. Exp. Med.* 192, 1069–1074.
- (39) Bowdish, D. M. E., Davidson, D. J., Scott, M. G., and Hancock, R. E. W. (2005) Immunomodulatory Activities of Small Host Defense Peptides. *Antimicrob. Agents Chemother.* 49, 1727–1732.
- (40) Kurosaka, K., Chen, Q., Yarovinsky, F., Oppenheim, J. J., and Yang, D. (2005) Mouse Cathelin-Related Antimicrobial Peptide Chemoattracts Leukocytes Using Formyl Peptide Receptor-Like 1/ Mouse Formyl Peptide Receptor-Like 2 as the Receptor and Acts as an Immune Adjuvant. *J. Immunol.* 174, 6257–6265.
- (41) Ohgami, K., Ilieva, I. B., Shiratori, K., Isogai, E., Yoshida, K., Kotake, S., Nishida, T., Mizuki, N., and Ohno, S. (2003) Effect of Human Cationic Antimicrobial Protein 18 Peptide on Endotoxin-Induced Uveitis in Rats. *Invest. Ophthalmol. Visual Sci.* 44, 4412–4418.
- (42) Stenger, S., Hanson, D. A., Teitelbaum, R., Dewan, P., Niazi, K. R., Froelich, C. J., Ganz, T., Thoma-Uszynski, S., Melián, A., Bogdan,

C., et al. (1998) An Antimicrobial Activity of Cytolytic T Cells Mediated by Granulysin. *Science* 282, 121–125.

(43) Wei, H.-M., Lin, L.-C., Wang, C.-F., Lee, Y.-J., Chen, Y.-T., and Liao, Y.-D. (2016) Antimicrobial Properties of an Immunomodulator - 15 kDa Human Granulysin. *PLoS One* 11, e0156321.

(44) Deng, A., Chen, S., Li, Q., Lyu, S.-c., Clayberger, C., and Krensky, A. M. (2005) Granulysin, a Cytolytic Molecule, Is Also a Chemoattractant and Proinflammatory Activator. *J. Immunol.* 174, 5243–5248.

(45) Ellsner, A., Duncan, M., Gavrilin, M., and Wewers, M. D. (2004) A Novel P2X<sub>7</sub> Receptor Activator, the Human Cathelicidin-Derived Peptide LL37, Induces IL-1 $\beta$  Processing and Release. *J. Immunol.* 172, 4987–4994.

(46) Braff, M. H., Hawkins, M. i. A., Nardo, A. D., Lopez-Garcia, B., Howell, M. D., Wong, C., Lin, K., Streib, J. E., Dorschner, R., Leung, D. Y. M., et al. (2005) Structure-Function Relationships among Human Cathelicidin Peptides: Dissociation of Antimicrobial Properties from Host Immunostimulatory Activities. *J. Immunol.* 174, 4271–4278.

(47) Yu, J., Mookherjee, N., Wee, K., Bowdish, D. M. E., Pistolic, J., Li, Y., Rehaume, L., and Hancock, R. E. W. (2007) Host Defense Peptide LL-37, in Synergy with Inflammatory Mediator IL-1 $\beta$ , Augments Immune Responses by Multiple Pathways. *J. Immunol.* 179, 7684–7691.

(48) Brown, K. L., and Hancock, R. E. W. (2006) Cationic host defense (antimicrobial) peptides. *Curr. Opin. Immunol.* 18, 24–30.

(49) Niyonsaba, F., Ushio, H., Nagaoka, I., Okumura, K., and Ogawa, H. (2005) The Human  $\beta$ -Defensins (-1, -2, -3, -4) and Cathelicidin LL-37 Induce IL-18 Secretion through p38 and ERK MAPK Activation in Primary Human Keratinocytes. *J. Immunol.* 175, 1776–1784.

(50) Niyonsaba, F., Ushio, H., Nakano, N., Ng, W., Sayama, K., Hashimoto, K., Nagaoka, I., Okumura, K., and Ogawa, H. (2007) Antimicrobial Peptides Human  $\beta$ -Defensins Stimulate Epidermal Keratinocyte Migration, Proliferation and Production of Proinflammatory Cytokines and Chemokines. *J. Invest. Dermatol.* 127, 594–604.

(51) Mookherjee, N., Brown, K. L., Bowdish, D. M. E., Doria, S., Falsafi, R., Hokamp, K., Roche, F. M., Mu, R., Doho, G. H., Pistolic, J., et al. (2006) Modulation of the TLR-Mediated Inflammatory Response by the Endogenous Human Host Defense Peptide LL-37. *J. Immunol.* 176, 2455–2464.

(52) Mookherjee, N., Wilson, H. L., Doria, S., Popowich, Y., Falsafi, R., Yu, J., Li, Y., Veatch, S., Roche, F. M., Brown, K. L., et al. (2006) Bovine and human cathelicidin cationic host defense peptides similarly suppress transcriptional responses to bacterial lipopolysaccharide. *J. Leukocyte Biol.* 80, 1563–1574.

(53) Rosenfeld, Y., Papo, N., and Shai, Y. (2006) Endotoxin (Lipopolysaccharide) Neutralization by Innate Immunity Host-Defense Peptides: Peptide Properties and Plausible Modes of Action. *J. Biol. Chem.* 281, 1636–1643.

(54) Tydell, C. C., Yuan, J., Tran, P., and Selsted, M. E. (2006) Bovine Peptidoglycan Recognition Protein-S: Antimicrobial Activity, Localization, Secretion, and Binding Properties. *J. Immunol.* 176, 1154–1162.

(55) Yibin, G., Jiang, Z., Hong, Z., Gengfa, L., Liangxi, W., Guo, W., and Yongling, L. (2005) A synthesized cationic tetradecapeptide from hornet venom kills bacteria and neutralizes lipopolysaccharide in vivo and in vitro. *Biochem. Pharmacol.* 70, 209–219.

(56) Chen, X., Dings, R. P. M., Nesselova, I., Debbert, S., Haseman, J. R., Maxwell, J., Hoye, T. R., and Mayo, K. H. (2006) Topomimetics of Amphipathic  $\beta$ -Sheet and Helix-Forming Bactericidal Peptides Neutralize Lipopolysaccharide Endotoxins. *J. Med. Chem.* 49, 7754–7765.

(57) Scott, M. G., Vreugdenhil, A. C. E., Buurman, W. A., Hancock, R. E. W., and Gold, M. R. (2000) Cutting Edge: Cationic Antimicrobial Peptides Block the Binding of Lipopolysaccharide (LPS) to LPS Binding Protein. *J. Immunol.* 164, 549–553.

(58) Di Nardo, A., Braff, M. H., Taylor, K. R., Na, C., Granstein, R. D., McInturff, J. E., Krutzik, S., Modlin, R. L., and Gallo, R. L. (2007)

Cathelicidin Antimicrobial Peptides Block Dendritic Cell TLR4 Activation and Allergic Contact Sensitization. *J. Immunol.* 178, 1829–1834.

(59) Nijnik, A., Pistolic, J., Filewod, N. C. J., and Hancock, R. E. W. (2012) Signaling Pathways Mediating Chemokine Induction in Keratinocytes by Cathelicidin LL-37 and Flagellin. *J. Innate Immun.* 4, 377–386.

(60) Lande, R., Gregorio, J., Facchinetti, V., Chatterjee, B., Wang, Y.-H., Homey, B., Cao, W., Wang, Y.-H., Su, B., Nestle, F. O., et al. (2007) Plasmacytoid dendritic cells sense self-DNA coupled with antimicrobial peptide. *Nature* 449, 564–569.

(61) Hurtado, P., and Au Peh, C. (2010) LL-37 Promotes Rapid Sensing of CpG Oligodeoxynucleotides by B Lymphocytes and Plasmacytoid Dendritic Cells. *J. Immunol.* 184, 1425–1435.

(62) Nakagawa, Y., and Gallo, R. L. (2015) Endogenous Intracellular Cathelicidin Enhances TLR9 Activation in Dendritic Cells and Macrophages. *J. Immunol.* 194, 1274–1284.

(63) Lai, Y., Adhikarakunnathu, S., Bhardwaj, K., Ranjith-Kumar, C. T., Wen, Y., Jordan, J. L., Wu, L. H., Dragnea, B., Mateo, L. S., and Kao, C. C. (2011) LL37 and Cationic Peptides Enhance TLR3 Signaling by Viral Double-stranded RNAs. *PLoS One* 6, e26632.

(64) Lee, E. Y., Takahashi, T., Curk, T., Dobnikar, J., Gallo, R. L., and Wong, G. C. L. (2017) Crystallinity of Double-Stranded RNA-Antimicrobial Peptide Complexes Modulates Toll-Like Receptor 3-Mediated Inflammation. *ACS Nano* 11, 12145–12155.

(65) Ganguly, D., Chamilos, G., Lande, R., Gregorio, J., Meller, S., Facchinetti, V., Homey, B., Barrat, F. J., Zal, T., and Gilliet, M. (2009) Self-RNA-antimicrobial peptide complexes activate human dendritic cells through TLR7 and TLR8. *J. Exp. Med.* 206, 1983–1994.

(66) Schmidt, N. W., Jin, F., Lande, R., Curk, T., Xian, W., Lee, C., Frasca, L., Frenkel, D., Dobnikar, J., Gilliet, M., et al. (2015) Liquid-crystalline ordering of antimicrobial peptide-DNA complexes controls TLR9 activation. *Nat. Mater.* 14, 696–700.

(67) Lee, E. Y., Lee, C. K., Schmidt, N. W., Jin, F., Lande, R., Curk, T., Frenkel, D., Dobnikar, J., Gilliet, M., and Wong, G. C. L. (2016) A review of immune amplification via ligand clustering by self-assembled liquid-crystalline DNA complexes. *Adv. Colloid Interface Sci.* 232, 17–24.

(68) Lee, E. Y., Lee, M. W., and Wong, G. C. L. (2018) Modulation of toll-like receptor signaling by antimicrobial peptides. *Semin. Cell Dev. Biol.*, DOI: DOI: 10.1016/j.semcdb.2018.02.002.

(69) Yount, N. Y., and Yeaman, M. R. (2004) Multidimensional signatures in antimicrobial peptides. *Proc. Natl. Acad. Sci. U. S. A.* 101, 7363–7368.

(70) Yount, N. Y., Gank, K. D., Xiong, Y. Q., Bayer, A. S., Pender, T., Welch, W. H., and Yeaman, M. R. (2004) Platelet Microbicidal Protein 1: Structural Themes of a Multifunctional Antimicrobial Peptide. *Antimicrob. Agents Chemother.* 48, 4395–4404.

(71) Yount, N. Y., Waring, A. J., Gank, K. D., Welch, W. H., Kupferwasser, D., and Yeaman, M. R. (2007) Structural correlates of antimicrobial efficacy in IL-8 and related human kinocidins. *Biochim. Biophys. Acta, Biomembr.* 1768, 598–608.

(72) Yeaman, M. R., and Yount, N. Y. (2007) Unifying themes in host defence effector polypeptides. *Nat. Rev. Microbiol.* 5, 727–740.

(73) Yount, N. Y., Kupferwasser, D., Spisni, A., Dutz, S. M., Ramjan, Z. H., Sharma, S., Waring, A. J., and Yeaman, M. R. (2009) Selective reciprocity in antimicrobial activity versus cytotoxicity of hBD-2 and crotamine. *Proc. Natl. Acad. Sci. U. S. A.* 106, 14972–14977.

(74) Yount, N. Y., and Yeaman, M. R. (2006) Structural congruence among membrane-active host defense polypeptides of diverse phylogeny. *Biochim. Biophys. Acta, Biomembr.* 1758, 1373–1386.

(75) Nguyen, L., and Vogel, H. (2012) Structural perspectives on antimicrobial chemokines. *Front. Immunol.* 3, 384.

(76) Yeaman, M. R., Yount, N. Y., Waring, A. J., Gank, K. D., Kupferwasser, D., Wiese, R., Bayer, A. S., and Welch, W. H. (2007) Modular determinants of antimicrobial activity in platelet factor-4 family kinocidins. *Biochim. Biophys. Acta, Biomembr.* 1768, 609–619.



- (77) Tang, Y.-Q., Yeaman, M. R., and Selsted, M. E. (2002) Antimicrobial Peptides from Human Platelets. *Infect. Immun.* 70, 6524–6533.
- (78) Cole, A. M., Ganz, T., Liese, A. M., Burdick, M. D., Liu, L., and Strieter, R. M. (2001) Cutting Edge: IFN-Inducible ELR<sup>-</sup> CXC Chemokines Display Defensin-Like Antimicrobial Activity. *J. Immunol.* 167, 623–627.
- (79) Björstad, Å., Fu, H., Karlsson, A., Dahlgren, C., and Bylund, J. (2005) Interleukin-8-Derived Peptide Has Antibacterial Activity. *Antimicrob. Agents Chemother.* 49, 3889–3895.
- (80) Meller, S., Di Domizio, J., Voo, K. S., Friedrich, H. C., Chamilos, G., Ganguly, D., Conrad, C., Gregorio, J., Le Roy, D., Roger, T., et al. (2015) T<sub>H</sub>17 cells promote microbial killing and innate immune sensing of DNA via interleukin 26. *Nat. Immunol.* 16, 970–979.
- (81) Poli, C., Augusto, J. F., Dauvé, J., Adam, C., Preisser, L., Larochette, V., Pignon, P., Savina, A., Blanchard, S., Subra, J. F., et al. (2017) IL-26 Confers Proinflammatory Properties to Extracellular DNA. *J. Immunol.* 198, 3650–3661.
- (82) Plataniias, L. C. (2005) Mechanisms of type-I- and type-II-interferon-mediated signalling. *Nat. Rev. Immunol.* 5, 375–386.
- (83) Kaplan, A., Lee, M. W., Wolf, A. J., Limon, J. J., Becker, C. A., Ding, M., Murali, R., Lee, E. Y., Liu, G. Y., Wong, G. C. L., et al. (2017) Direct Antimicrobial Activity of IFN- $\beta$ . *J. Immunol.* 198, 4036–4045.
- (84) Boxx, G. M., and Cheng, G. (2016) The Roles of Type I Interferon in Bacterial Infection. *Cell Host Microbe* 19, 760–769.
- (85) Kovarik, P., Castiglia, V., Ivin, M., and Ebner, F. (2016) Type I Interferons in Bacterial Infections: A Balancing Act. *Front. Immunol.* 7, 652.
- (86) Decker, T., Müller, M., and Stockinger, S. (2005) The Yin and Yang of type I interferon activity in bacterial infection. *Nat. Rev. Immunol.* 5, 675–687.
- (87) Donato, R., Cannon, B. R., Sorci, G., Riuizi, F., Hsu, K., Weber, D. J., and Geczy, C. L. (2013) Functions of S100 Proteins. *Curr. Mol. Med.* 13, 24–57.
- (88) Elsbach, P. (2003) What is the real role of antimicrobial polypeptides that can mediate several other inflammatory responses? *J. Clin. Invest.* 111, 1643–1645.
- (89) Eckert, R. L., Broome, A.-M., Ruse, M., Robinson, N., Ryan, D., and Lee, K. (2004) S100 Proteins in the Epidermis. *J. Invest. Dermatol.* 123, 23–33.
- (90) Donato, R. (2001) S100: a multigenic family of calcium-modulated proteins of the EF-hand type with intracellular and extracellular functional roles. *Int. J. Biochem. Cell Biol.* 33, 637–668.
- (91) Vogl, T., Tenbrock, K., Ludwig, S., Leukert, N., Ehrhardt, C., van Zoelen, M. A. D., Nacken, W., Foell, D., van der Poll, T., Sorg, C., et al. (2007) Mrp8 and Mrp14 are endogenous activators of Toll-like receptor 4, promoting lethal, endotoxin-induced shock. *Nat. Med.* 13, 1042–1049.
- (92) Realegeno, S., Kelly-Scumpia, K. M., Dang, A. T., Lu, J., Teles, R., Liu, P. T., Schenk, M., Lee, E. Y., Schmidt, N. W., Wong, G. C. L., et al. (2016) S100A12 Is Part of the Antimicrobial Network against *Mycobacterium leprae* in Human Macrophages. *PLoS Pathog.* 12, e1005705.
- (93) Yang, Z., Tao, T., Raftery, M. J., Youssef, P., Di Girolamo, N., and Geczy, C. L. (2001) Proinflammatory properties of the human S100 protein S100A12. *J. Leukocyte Biol.* 69, 986–994.
- (94) Michalek, M., Gelhaus, C., Hecht, O., Podschun, R., Schröder, J. M., Leippe, M., and Grötzinger, J. (2009) The human antimicrobial protein psoriasin acts by permeabilization of bacterial membranes. *Dev. Comp. Immunol.* 33, 740–746.
- (95) Raines, R. T. (1998) Ribonuclease A. *Chem. Rev.* 98, 1045–1066.
- (96) Boix, E., and Nogue, M. V. (2007) Mammalian antimicrobial proteins and peptides: overview on the RNase A superfamily members involved in innate host defence. *Mol. Biosyst.* 3, 317–335.
- (97) Rosenberg, H. F. (1995) Recombinant Human Eosinophil Cationic Protein: Ribonuclease Activity Is Not Essential for Cytotoxicity. *J. Biol. Chem.* 270, 7876–7881.
- (98) Huang, Y.-C., Lin, Y.-M., Chang, T.-W., Wu, S.-J., Lee, Y.-S., Chang, M. D.-T., Chen, C., Wu, S.-H., and Liao, Y.-D. (2007) The Flexible and Clustered Lysine Residues of Human Ribonuclease 7 Are Critical for Membrane Permeability and Antimicrobial Activity. *J. Biol. Chem.* 282, 4626–4633.
- (99) Köten, B., Simanski, M., Gläser, R., Podschun, R., Schröder, J.-M., and Harder, J. (2009) RNase 7 Contributes to the Cutaneous Defense against *Enterococcus faecium*. *PLoS One* 4, e6424.
- (100) Hogan, S. P., Rosenberg, H. F., Moqbel, R., Phipps, S., Foster, P. S., Lacy, P., Kay, A. B., and Rothenberg, M. E. (2008) Eosinophils: Biological Properties and Role in Health and Disease. *Clin. Exp. Allergy* 38, 709–750.
- (101) Gomes, I., Mathur, S. K., Espenshade, B. M., Mori, Y., Varga, J., and Ackerman, S. J. (2005) Eosinophil-fibroblast interactions induce fibroblast IL-6 secretion and extracellular matrix gene expression: Implications in fibrogenesis. *J. Allergy Clin. Immunol.* 116, 796–804.
- (102) Young, J. D.-E., Peterson, C. G. B., Venge, P., and Cohn, Z. A. (1986) Mechanism of membrane damage mediated by human eosinophil cationic protein. *Nature* 321, 613–616.
- (103) Lehrer, R. I., Szklarek, D., Barton, A., Ganz, T., Hamann, K. J., and Gleich, G. J. (1989) Antibacterial properties of eosinophil major basic protein and eosinophil cationic protein. *J. Immunol.* 142, 4428–4434.
- (104) Carreras, E., Boix, E., Rosenberg, H. F., Cuchillo, C. M., and Nogués, M. V. (2003) Both Aromatic and Cationic Residues Contribute to the Membrane-Lytic and Bactericidal Activity of Eosinophil Cationic Protein. *Biochemistry* 42, 6636–6644.
- (105) Torrent, M., de la Torre, B. G., Nogués, V. M., Andreu, D., and Boix, E. (2009) Bactericidal and membrane disruption activities of the eosinophil cationic protein are largely retained in an N-terminal fragment. *Biochem. J.* 421, 425–434.
- (106) Pulido, D., Torrent, M., Andreu, D., Nogués, M. V., and Boix, E. (2013) Two Human Host Defense Ribonucleases against Mycobacteria, the Eosinophil Cationic Protein (RNase 3) and RNase 7. *Antimicrob. Agents Chemother.* 57, 3797–3805.
- (107) Torrent, M., Cuyás, E., Carreras, E., Navarro, S., López, O., de la Maza, A., Nogués, M. V., Reshetnyak, Y. K., and Boix, E. (2007) Topography Studies on the Membrane Interaction Mechanism of the Eosinophil Cationic Protein. *Biochemistry* 46, 720–733.
- (108) Kato, M., Yamada, Y., Maruyama, K., and Hayashi, Y. (2010) Serum Eosinophil Cationic Protein and 27 Cytokines/Chemokines in Acute Exacerbation of Childhood Asthma. *Int. Arch. Allergy Immunol.* 152, 62–66.
- (109) Venge, Carlson, Karawaczyk, and Trulsson (1999) Eosinophil cationic protein (ECP): molecular and biological properties and the use of ECP as a marker of eosinophil activation in disease. *Clin. Exp. Allergy* 29, 1172–1186.
- (110) Harder, J., and Schröder, J.-M. (2002) RNase 7, a Novel Innate Immune Defense Antimicrobial Protein of Healthy Human Skin. *J. Biol. Chem.* 277, 46779–46784.
- (111) Spencer, J. D., Schwaderer, A. L., Wang, H., Bartz, J., Kline, J., Eichler, T., DeSouza, K. R., Sims-Lucas, S., Baker, P., and Hains, D. S. (2013) Ribonuclease 7, an antimicrobial peptide upregulated during infection, contributes to microbial defense of the human urinary tract. *Kidney Int.* 83, 615–625.
- (112) Schmidt, N. W., and Wong, G. C. L. (2013) Antimicrobial peptides and induced membrane curvature: Geometry, coordination chemistry, and molecular engineering. *Curr. Opin. Solid State Mater. Sci.* 17, 151–163.
- (113) Dathe, M., and Wieprecht, T. (1999) Structural features of helical antimicrobial peptides: their potential to modulate activity on model membranes and biological cells. *Biochim. Biophys. Acta, Biomembr.* 1462, 71–87.
- (114) Dathe, M., Nikolenko, H., Meyer, J., Beyermann, M., and Bienert, M. (2001) Optimization of the antimicrobial activity of magainin peptides by modification of charge. *FEBS Lett.* 501, 146–150.



- (115) Matsuzaki, K., Nakamura, A., Murase, O., Sugishita, K.-i., Fujii, N., and Miyajima, K. (1997) Modulation of Magainin 2–Lipid Bilayer Interactions by Peptide Charge. *Biochemistry* 36, 2104–2111.
- (116) Wieprecht, T., Dathe, M., Beyermann, M., Krause, E., Maloy, W. L., MacDonald, D. L., and Bienert, M. (1997) Peptide Hydrophobicity Controls the Activity and Selectivity of Magainin 2 Amide in Interaction with Membranes. *Biochemistry* 36, 6124–6132.
- (117) Kustanovich, I., Shalev, D. E., Mikhlin, M., Gaidukov, L., and Mor, A. (2002) Structural Requirements for Potent Versus Selective Cytotoxicity for Antimicrobial Dermaseptin S4 Derivatives. *J. Biol. Chem.* 277, 16941–16951.
- (118) Tytler, E. M., Segrest, J. P., Eband, R. M., Nie, S. Q., Eband, R. F., Mishra, V. K., Venkatachalapathi, Y. V., and Anantharamaiah, G. M. (1993) Reciprocal effects of apolipoprotein and lytic peptide analogs on membranes. Cross-sectional molecular shapes of amphipathic alpha helices control membrane stability. *J. Biol. Chem.* 268, 22112–22118.
- (119) Drin, G., and Antonny, B. (2010) Amphipathic helices and membrane curvature. *FEBS Lett.* 584, 1840–1847.
- (120) Eband, R. M., Shai, Y., Segrest, J. P., and Anantharamaiah, G. M. (1995) Mechanisms for the modulation of membrane bilayer properties by amphipathic helical peptides. *Biopolymers* 37, 319–338.
- (121) Segrest, J. P., De Loof, H., Dohlman, J. G., Brouillette, C. G., and Anantharamaiah, G. M. (1990) Amphipathic helix motif: Classes and properties. *Proteins: Struct., Funct., Genet.* 8, 103–117.
- (122) Wang, G., Li, X., and Wang, Z. (2016) APD3: the antimicrobial peptide database as a tool for research and education. *Nucleic Acids Res.* 44, D1087–D1093.
- (123) Schmidt, N. W., Mishra, A., Lai, G. H., Davis, M., Sanders, L. K., Tran, D., Garcia, A., Tai, K. P., McCray, P. B., Ouellette, A. J., et al. (2011) Criterion for Amino Acid Composition of Defensins and Antimicrobial Peptides Based on Geometry of Membrane Destabilization. *J. Am. Chem. Soc.* 133, 6720–6727.
- (124) Gelbart, W. M., Ben-Shaul, A., and Roux, D. (1994) *Micelles, Membranes, Microemulsions, and Monolayers*, p 608, Springer-Verlag, New York, NY.
- (125) Schmidt, N. W., Tai, K. P., Kamdar, K., Mishra, A., Lai, G. H., Zhao, K., Ouellette, A. J., and Wong, G. C. L. (2012) Arginine in  $\alpha$ -Defensins: Differential Effects on Bactericidal Activity Correspond to Geometry of Membrane Curvature Generation and Peptide–Lipid Phase Behavior. *J. Biol. Chem.* 287, 21866–21872.
- (126) Yang, L., Gordon, V. D., Trinkle, D. R., Schmidt, N. W., Davis, M. A., DeVries, C., Som, A., Cronan, J. E., Tew, G. N., and Wong, G. C. L. (2008) Mechanism of a prototypical synthetic membrane-active antimicrobial: Efficient hole-punching via interaction with negative intrinsic curvature lipids. *Proc. Natl. Acad. Sci. U. S. A.* 105, 20595–20600.
- (127) Hu, K., Schmidt, N. W., Zhu, R., Jiang, Y., Lai, G. H., Wei, G., Palermo, E. F., Kuroda, K., Wong, G. C. L., and Yang, L. (2013) A Critical Evaluation of Random Copolymer Mimesis of Homogeneous Antimicrobial Peptides. *Macromolecules* 46, 1908–1915.
- (128) Lee, M. W., Chakraborty, S., Schmidt, N. W., Murgai, R., Gellman, S. H., and Wong, G. C. L. (2014) Two interdependent mechanisms of antimicrobial activity allow for efficient killing in nylon-3-based polymeric mimics of innate immunity peptides. *Biochim. Biophys. Acta, Biomembr.* 1838, 2269–2279.
- (129) Xiong, M., Lee, M. W., Mansbach, R. A., Song, Z., Bao, Y., Peek, R. M., Yao, C., Chen, L.-F., Ferguson, A. L., Wong, G. C. L., et al. (2015) Helical antimicrobial polypeptides with radial amphiphilicity. *Proc. Natl. Acad. Sci. U. S. A.* 112, 13155–13160.
- (130) Schmidt, N. W., Lis, M., Zhao, K., Lai, G. H., Alexandrova, A. N., Tew, G. N., and Wong, G. C. L. (2012) Molecular Basis for Nanoscopic Membrane Curvature Generation from Quantum Mechanical Models and Synthetic Transporter Sequences. *J. Am. Chem. Soc.* 134, 19207–19216.
- (131) Mishra, A., Lai, G. H., Schmidt, N. W., Sun, V. Z., Rodriguez, A. R., Tong, R., Tang, L., Cheng, J., Deming, T. J., Kamei, D. T., et al. (2011) Translocation of HIV TAT peptide and analogues induced by multiplexed membrane and cytoskeletal interactions. *Proc. Natl. Acad. Sci. U. S. A.* 108, 16883–16888.
- (132) Schmidt, N., Mishra, A., Lai, G. H., and Wong, G. C. L. (2010) Arginine-rich cell-penetrating peptides. *FEBS Lett.* 584, 1806–1813.
- (133) Milletti, F. (2012) Cell-penetrating peptides: classes, origin, and current landscape. *Drug Discovery Today* 17, 850–860.
- (134) Bechara, C., and Sagan, S. (2013) Cell-penetrating peptides: 20 years later, where do we stand? *FEBS Lett.* 587, 1693–1702.
- (135) Frankel, A. D., and Pabo, C. O. (1988) Cellular uptake of the tat protein from human immunodeficiency virus. *Cell* 55, 1189–1193.
- (136) Ziegler, A., Nervi, P., Dürrenberger, M., and Seelig, J. (2005) The Cationic Cell-Penetrating Peptide CPP<sup>TAT</sup> Derived from the HIV-1 Protein TAT Is Rapidly Transported into Living Fibroblasts: Optical, Biophysical, and Metabolic Evidence. *Biochemistry* 44, 138–148.
- (137) Deshayes, S., Morris, M. C., Divita, G., and Heitz, F. (2006) Interactions of amphipathic CPPs with model membranes. *Biochim. Biophys. Acta, Biomembr.* 1758, 328–335.
- (138) Ziegler, A. (2008) Thermodynamic studies and binding mechanisms of cell-penetrating peptides with lipids and glycosaminoglycans. *Adv. Drug Delivery Rev.* 60, 580–597.
- (139) Ludtke, S. J., He, K., Heller, W. T., Harroun, T. A., Yang, L., and Huang, H. W. (1996) Membrane Pores Induced by Magainin. *Biochemistry* 35, 13723–13728.
- (140) Jiao, C.-Y., Delaroche, D., Burlina, F., Alves, I. D., Chassaing, G., and Sagan, S. (2009) Translocation and Endocytosis for Cell-penetrating Peptide Internalization. *J. Biol. Chem.* 284, 33957–33965.
- (141) Maiolo, J. R., Ferrer, M., and Ottinger, E. A. (2005) Effects of cargo molecules on the cellular uptake of arginine-rich cell-penetrating peptides. *Biochim. Biophys. Acta, Biomembr.* 1712, 161–172.
- (142) Iwasa, A., Akita, H., Khalil, I., Kogure, K., Futaki, S., and Harashima, H. (2006) Cellular uptake and subsequent intracellular trafficking of R8-liposomes introduced at low temperature. *Biochim. Biophys. Acta, Biomembr.* 1758, 713–720.
- (143) Ter-Avetisyan, G., Tünnemann, G., Nowak, D., Nitschke, M., Herrmann, A., Drab, M., and Cardoso, M. C. (2009) Cell Entry of Arginine-rich Peptides Is Independent of Endocytosis. *J. Biol. Chem.* 284, 3370–3378.
- (144) Ferrari, A., Pellegrini, V., Arcangeli, C., Fittipaldi, A., Giacca, M., and Beltram, F. (2003) Caveolae-Mediated internalization of extracellular HIV-1 tat fusion proteins visualized in real time. *Mol. Ther.* 8, 284–294.
- (145) Richard, J. P., Melikov, K., Brooks, H., Prevot, P., Lebleu, B., and Chernomordik, L. V. (2005) Cellular Uptake of Unconjugated TAT Peptide Involves Clathrin-dependent Endocytosis and Heparan Sulfate Receptors. *J. Biol. Chem.* 280, 15300–15306.
- (146) Lee, M. W., Han, M., Bossa, G. V., Snell, C., Song, Z., Tang, H., Yin, L., Cheng, J., May, S., Luijten, E., et al. (2017) Interactions between Membranes and “Metaphilic” Polypeptide Architectures with Diverse Side-Chain Populations. *ACS Nano* 11, 2858–2871.
- (147) Schmidt, N. W., Mishra, A., Wang, J., DeGrado, W. F., and Wong, G. C. L. (2013) Influenza Virus A M2 Protein Generates Negative Gaussian Membrane Curvature Necessary for Budding and Scission. *J. Am. Chem. Soc.* 135, 13710–13719.
- (148) Yao, H., Lee, M. W., Waring, A. J., Wong, G. C. L., and Hong, M. (2015) Viral fusion protein transmembrane domain adopts  $\beta$ -strand structure to facilitate membrane topological changes for virus–cell fusion. *Proc. Natl. Acad. Sci. U. S. A.* 112, 10926–10931.
- (149) Cunden, L. S., Gaillard, A., and Nolan, E. M. (2016) Calcium ions tune the zinc-sequestering properties and antimicrobial activity of human S100A12. *Chem. Sci.* 7, 1338–1348.
- (150) Tossi, A., Sandri, L., and Giangaspero, A. (2000) Amphipathic,  $\alpha$ -helical antimicrobial peptides. *Biopolymers* 55, 4–30.
- (151) Fjell, C. D., Jenssen, H., Hilpert, K., Cheung, W. A., Panté, N., Hancock, R. E. W., and Cherkasov, A. (2009) Identification of Novel Antibacterial Peptides by Chemoinformatics and Machine Learning. *J. Med. Chem.* 52, 2006–2015.
- (152) Porto, W. F., Fernandes, F. C., and Franco, O. L. (2010) In *Advances in Bioinformatics and Computational Biology* (Ferreira, C. E., Miyano, S., and Stadler, P. F., Eds.) pp 59–62, Springer, Berlin,

Heidelberg, 5th Brazilian Symposium on Bioinformatics, Rio de Janeiro, Brazil, August 31–September 3, 2010.

(153) Porto, W. F., Pires, A. S., and Franco, O. L. (2012) CS-AMPPred: An Updated SVM Model for Antimicrobial Activity Prediction in Cysteine-Stabilized Peptides. *PLoS One* 7, e51444.

(154) Hilpert, K., Fjell, C. D., and Cherkasov, A. (2008) Short Linear Cationic Antimicrobial Peptides: Screening, Optimizing, and Prediction. *Peptide-Based Drug Design* (Otvos, L., Ed.) pp 127–159, Humana Press, Totowa, NJ.

(155) Lata, S., Sharma, B., and Raghava, G. (2007) Analysis and prediction of antibacterial peptides. *BMC Bioinf.* 8, 263.

(156) Lee, E. Y., Fulan, B. M., Wong, G. C. L., and Ferguson, A. L. (2016) Mapping membrane activity in undiscovered peptide sequence space using machine learning. *Proc. Natl. Acad. Sci. U. S. A.* 113, 13588–13593.

(157) Lee, E. Y., Wong, G. C. L., and Ferguson, A. L. (2018) Machine learning-enabled discovery and design of membrane-active peptides. *Bioorg. Med. Chem.* 26, 2708.

(158) Lee, E. Y., Lee, M. W., Fulan, B. M., Ferguson, A. L., and Wong, G. C. L. (2017) What can machine learning do for antimicrobial peptides, and what can antimicrobial peptides do for machine learning? *Interface Focus* 7, 20160153.

(159) Wang, Z., and Wang, G. (2004) APD: the Antimicrobial Peptide Database. *Nucleic Acids Res.* 32, D590–D592.

(160) Wang, G., Li, X., and Wang, Z. (2009) APD2: the updated antimicrobial peptide database and its application in peptide design. *Nucleic Acids Res.* 37, D933–D937.

(161) Kozma, D., Simon, I., and Tuszáný, G. E. (2012) PDBTM: Protein Data Bank of transmembrane proteins after 8 years. *Nucleic Acids Res.* 41, D524–D529.

(162) Lee, M. W., Lee, E. Y., Lai, G. H., Kennedy, N. W., Posey, A. E., Xian, W., Ferguson, A. L., Hill, R. B., and Wong, G. C. L. (2017) Molecular Motor Dnm1 Synergistically Induces Membrane Curvature To Facilitate Mitochondrial Fission. *ACS Cent. Sci.* 3, 1156–1167.

(163) Bleazard, W., McCaffery, J. M., King, E. J., Bale, S., Mozdy, A., Tieu, Q., Nunnari, J., and Shaw, J. M. (1999) The dynamin-related GTPase Dnm1 regulates mitochondrial fission in yeast. *Nat. Cell Biol.* 1, 298–304.

(164) Carlton, J. G., and Martin-Serrano, J. (2009) The ESCRT machinery: new functions in viral and cellular biology. *Biochem. Soc. Trans.* 37, 195–199.

(165) Chen, B. J., and Lamb, R. A. (2008) Mechanisms for enveloped virus budding: Can some viruses do without an ESCRT? *Virology* 372, 221–232.

(166) Pornillos, O., Garrus, J. E., and Sundquist, W. I. (2002) Mechanisms of enveloped RNA virus budding. *Trends Cell Biol.* 12, 569–579.

(167) Bruce, E. A., Medcalf, L., Crump, C. M., Noton, S. L., Stuart, A. D., Wise, H. M., Elton, D., Bowers, K., and Digard, P. (2009) Budding of filamentous and non-filamentous influenza A virus occurs via a VPS4 and VPS28-independent pathway. *Virology* 390, 268–278.

(168) Stewart, S. M., and Pekosz, A. (2011) Mutations in the Membrane-Proximal Region of the Influenza A Virus M2 Protein Cytoplasmic Tail Have Modest Effects on Virus Replication. *J. Virol.* 85, 12179–12187.

(169) Schnell, J. R., and Chou, J. J. (2008) Structure and mechanism of the M2 proton channel of influenza A virus. *Nature* 451, 591–595.

(170) Takeda, M., Pekosz, A., Shuck, K., Pinto, L. H., and Lamb, R. A. (2002) Influenza A Virus M2 Ion Channel Activity Is Essential for Efficient Replication in Tissue Culture. *J. Virol.* 76, 1391–1399.

(171) Watanabe, T., Watanabe, S., Ito, H., Kida, H., and Kawaoka, Y. (2001) Influenza A Virus Can Undergo Multiple Cycles of Replication without M2 Ion Channel Activity. *J. Virol.* 75, S656–S662.

(172) Iwatsuki-Horimoto, K., Horimoto, T., Noda, T., Kiso, M., Maeda, J., Watanabe, S., Muramoto, Y., Fujii, K., and Kawaoka, Y. (2006) The Cytoplasmic Tail of the Influenza A Virus M2 Protein Plays a Role in Viral Assembly. *J. Virol.* 80, 5233–5240.

(173) Rossman, J. S., Jing, X., Leser, G. P., and Lamb, R. A. (2010) Influenza Virus M2 Protein Mediates ESCRT-Independent Membrane Scission. *Cell* 142, 902–913.

(174) Rossman, J. S., Jing, X., Leser, G. P., Balannik, V., Pinto, L. H., and Lamb, R. A. (2010) Influenza Virus M2 Ion Channel Protein Is Necessary for Filamentous Virion Formation. *J. Virol.* 84, 5078–5088.

(175) Rossman, J. S., and Lamb, R. A. (2011) Influenza virus assembly and budding. *Virology* 411, 229–236.

(176) Lewis, K. (2007) Persister cells, dormancy and infectious disease. *Nat. Rev. Microbiol.* 5, 48–56.

(177) Keren, I., Shah, D., Spoering, A., Kaldalu, N., and Lewis, K. (2004) Specialized Persister Cells and the Mechanism of Multidrug Tolerance in *Escherichia coli*. *J. Bacteriol.* 186, 8172–8180.

(178) Spoering, A. L., and Lewis, K. (2001) Biofilms and Planktonic Cells of *Pseudomonas aeruginosa* Have Similar Resistance to Killing by Antimicrobials. *J. Bacteriol.* 183, 6746–6751.

(179) Schmidt, N. W., Deshayes, S., Hawker, S., Blacker, A., Kasko, A. M., and Wong, G. C. L. (2014) Engineering Persister-Specific Antibiotics with Synergistic Antimicrobial Functions. *ACS Nano* 8, 8786–8793.

(180) Schmidt, N. W., Agak, G. W., Deshayes, S., Yu, Y., Blacker, A., Champer, J., Xian, W., Kasko, A. M., Kim, J., and Wong, G. C. L. (2015) Pentobra: A Potent Antibiotic with Multiple Layers of Selective Antimicrobial Mechanisms against *Propionibacterium Acnes*. *J. Invest. Dermatol.* 135, 1581–1589.

(181) Deshayes, S., Xian, W., Schmidt, N. W., Kordbacheh, S., Lieng, J., Wang, J., Zamer, S., Germain, S. S., Voyer, L., Thulin, J., et al. (2017) Designing Hybrid Antibiotic Peptide Conjugates To Cross Bacterial Membranes. *Bioconjugate Chem.* 28, 793–804.

(182) Derossi, D., Chassaing, G., and Prochiantz, A. (1998) Trojan peptides: the penetratin system for intracellular delivery. *Trends Cell Biol.* 8, 84–87.

(183) Diener, C., Garza Ramos Martínez, G., Moreno Blas, D., Castillo González, D. A., Corzo, G., Castro-Oregon, S., and Del Rio, G. (2016) Effective Design of Multifunctional Peptides by Combining Compatible Functions. *PLoS Comput. Biol.* 12, e1004786.